

Structural Characterization of Histone Deacetylase from *Plasmodium Falciparum*

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Abstract – Histone deacetylase (HDAC) is the key enzyme responsible for epigenetic regulation of an organism. This protein has been involved in transcriptional regulation of many proteins associated with chromatin remodelling. Homologs of histone deacetylase are also found in malaria parasite *Plasmodium falciparum* where it plays major role in regulation of key pathways of parasite. In this study, we determined the three-dimensional structure of histone deacetylase from *Plasmodium falciparum* (PfHDAC) by using homology modelling tools available at Swiss Modeller server and Modweb. Modelled structure was validated using Ramachandran plot and active site determination was performed using CASTp. We believe that structural analysis of PfHDAC could be pivotal in discovering new drug like molecules against malaria parasite.

Keywords: HDAC, Transcription regulation, molecular modelling, malaria, drug discovery

I. INTRODUCTION

Malaria is one of the major problems in many developing countries which are caused by the protozoan parasite *Plasmodium*. Several cases are reported annually. Many drugs have been invented against malaria but developing resistant in malaria parasite has raised the concern of identifying new protein molecules which can be treated as viable drug target. There are many pathways crucial for parasite survival and some of them are very unique to *Plasmodium*. Regulation of transcription remains the major pathway in survival of any organism including malaria parasite where post-translational modification of histone proteins are very critical. Histone acetyl transferase (HAT) and histone deacetylase are two major enzymes involved in transcriptional regulation [1]. HAT catalyzes acetylation on histone lysine residue whereas HDAC does the removal of acetyl group from histone leads to chromatin condensation and transcriptional repression. Histone deacetylases (HDACs) is the enzyme generally localized in the nucleus [2]. HDACs are classified into various classes and sub-classes based on their catalytic centre [3]. Several studies have shown that HDACs play crucial role in cell survival and proliferation [4]. Many other proteins along

with the histones, which are involved in the cell migration, cell proliferation and cell death, are target of HDACs. When interact with histones, these proteins catalyze the deacetylation of α -acetyl lysine at the N-terminal of histone core. Inhibition of HDACs activity has been established as a proven cancer therapy [5].

Malaria parasite *Plasmodium* harbour many histone deacetylases (PfHDACs), which have been shown to regulate the transcription of many genes. Inhibition of PfHDAC resulted in the change of expression profile of many genes from all the developmental stages of asexual life cycle of parasite viz. ring, trophozoite and schizont [6]. The level of acetylation and chromatin structure was also altered. Though, biochemical data on PfHDACs are available, structural characterization remain obscure. Hence, in this study, we evaluated the structural properties of PfHDAC using in-silico structure determination method and dissected the structure with identification of active site. Our studies provided the structural framework of PfHDAC in three-dimension space which can be utilized for high-through put drug screening against malaria parasite.

II. MATERIALS AND METHODS

The sequence of PfHDAC was retrieved from PlasmoDB (PFI1260c). 3MAX pdb structure was used as a template for homology modeling. Identification of template structures was carried out using NCBI BlastP. Modeller [7]. and Swiss Model Server were used to build the in-silico structure of PfHDAC. Structure validation was performed with Ramachandran plot using online server RAMPAGE [8]. Modelled structure of PfHDAC was submitted to CASTp [9]. for active site prediction. Figures and images were developed using CHIMERA [10].

III. RESULTS AND DISCUSSION

The modelled structure of PfHDAC is shown in Figure 1 with ribbon diagram along with surface topology representation. Structure of PfHDAC is broadly divided into rossman like fold and zinc binding fold. Bound zinc residues

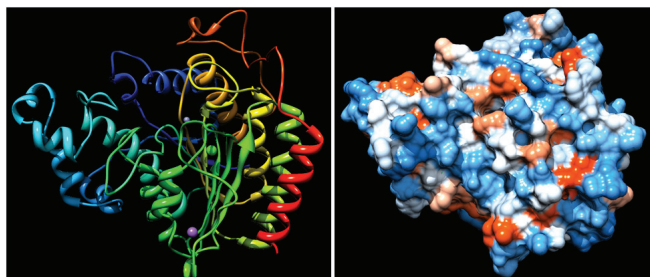


Fig. 1 : Modelled structure of PfHDAC. A) Ribbon diagram; B) Surface topology

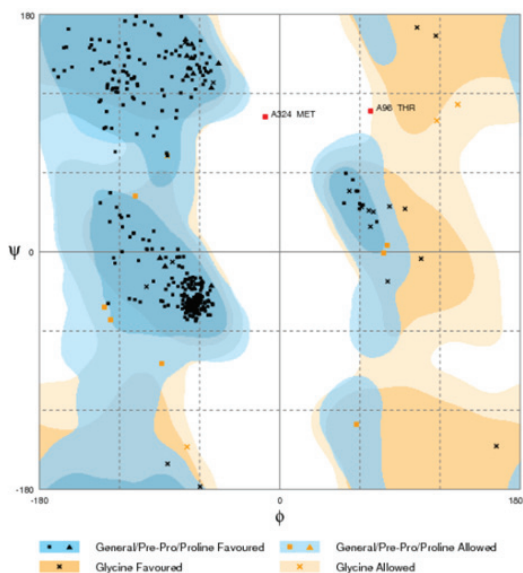


Fig. 2 : Ramachandran plot PfHDAC using RAMPAGE

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+ RKKVAFHDP D1 GSYV2 GAG3 H4 P5 K6 P7 Q8 R9 T10 R11 M12 T13 H14 S15 L16 I17 V18 S19 Y20 N21 L22 Y23 K24 Y25 M26 E27 V28 Y29 R30 P31
* KSDVNELTLF HDYEYIDFLS S32 I33 S34 L35 E36 N37 Y38 R39 E40 F41 T42 Y43 Q44 L45 K46 R47 F48 N49 V50 G51 E52 A53 T54 D55 C56 P57 A58 F59 D60 G61
# L62 F63 Q64 R65 Q66 Q67 S68 E69 G70 A71 S72 I73 D74 G75 A76 S77 K78 E79 L80 N81 H82 C83 A84 D85 I86 C87 V88 N89 W90 S91 G92 G93 L94 H95 H96 A97 K98 M99 S100 E101 A102 S103 G104 F105 C106 V107 I108 N109 D110
# I111 V112 L113 G114 I115 L116 L117 K118 Y119 H120 A121 R122 V123 M124 Y125 I126 D127 I128 D129 V130 E131 H132 G133 D134 G135 V136 E137 E138 A139 F140 Y141 V142 T143 H144 R145 V146 M147 T148 V149 S150 F151 H152 K153 F154 C155 D156 Y157 F158
# P159 G160 T161 G162 D163 I164 T165 D166 V167 G168 V169 N170 H171 G172 K173 Y174 S175 V176 N177 V178 P179 L180 N181 D182 G183 M184 T185 D186 D187 A188 F189 V190 D191 L192 F193 K194 V195 V196 I197 D198 E199 C200 V201 Q202 T203 Y204 R205 P206 G207
# A208 I209 I210 I211 C212 G213 C214 A215 D216 S217 L218 T219 G220 D221 R222 L223 G224 R225 F226 N227 L228 T229 I230 K231 G232 H233 A234 R235 C236 V237 E238 H239 V240 S241 Y242 N243 I244 P245 L246 V247 L248 G249 G250 G251 Y252 T253 I254
# R255 N256 V257 S258 R259 C260 W261 A262 Y263 E264 T265 G266 V267 V268 L269 N270 K271 H272 H273 E274 M275 P276 D277 Q278 I279 S280 L281 N282 D283 Y284 Y285 A286 P287 D288 F289 Q290 L291 H292 L293 Q294 S295 N296 I297 P298 N299 Y300
# N301 S302 P303 E304 H305 L306 S307 R308 I309 K310 M311 E312 I313 A314 E315 N316 L317 R318
    
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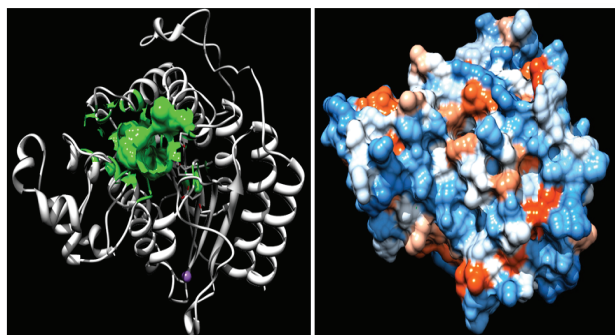


Figure 3: Prediction of active site of PfHDAC using CASTp. A) Active site prediction using CASTp where active site residues shown in green colour in between amino acid sequence of protein; B) & C) Position of active site residues are shown in three-dimensional space in ribbon form and surface topology respectively using CHIMERA.

are also shown in structure. Modelled PfHDAC structure is helical dominant with intermittent loops are hanging out and middle part of the structure is mostly occupied by beta sheets. Surface topology diagram shows the patches of negatively charged residues all over the surface probably required for interaction with positively charged histone proteins.

A large cavity is predicted to be active site of the enzyme PfHDAC with the cavity size of volume 409 and area of 401 angstrom. The cavity is shown with the green color in the diagram (Fig. 3). A zinc residue is also seen near active site, helps in the catalysis of the enzyme. Overall the compact active site with zinc residue perfectly setup the platform for deacetylation to occur. In addition, structure validation was done by using Ramachandran plot which shows that most of the residues are in favoured and allowed region to prove the authenticity of homology modeling (Fig. 2).

IV. CONCLUSION

Biochemical characterization of HDAC from Plasmodium falciparum has been done but structural information was missing. This lack of information clearly blocks the possibility of transferring available facts of transcription regulation for development of new anti-malarial drugs. Thus, an in-silico approach is the most efficient way of structural characterization of proteins. Molecular modeling of the PfHDAC provided us the 3D structures of the protein. Three-dimension structure of the parasite protein could act as a starting material for the in-silico drug screening. Not only that, but the prediction of the active site might also be useful in understanding the enzymatic activity of the protein which is crucial in deciphering the regulation of transcriptional control. In addition, modelled PfHDAC can be compared with its human counterparts for structural discrepancy, which could also fasten the process of drug development against malaria parasites.

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REFERENCES

- [1] Li J, Lin Q, Wang W, Wade P, Wong J, "Specific targeting and constitutive association of histone deacetylase complexes during transcriptional repression", *Genes Dev*, 2002, Vol. 16, pp. 687-692.
- [2] Dokmanovic M., Marks P.A., "Prospects: histone deacetylase inhibitors", *J Cell Biochem*, 2005, Vol. 96, pp.293-304.
- [3] Lagger G., O'Carroll D., Rembold M., et al, "Essential function of histone deacetylase 1 in proliferation control and CDK inhibitor repression", *EMBO J*, 2002, Vol. 21, pp.2672-81.
- [4] Xu W., Parmigiani R., PA M., "Histone deacetylase inhibitors: molecular mechanism of action", *Oncogene*, 2007, Vol. 26, pp. 5541-52.
- [5] Bolden J.E., Peart M.J., Johnstone R.W., "Anticancer activities of histone deacetylase inhibitors", *Nat Rev Drug Discov*, 2006, Vol. 5, pp.769-84.
- [6] Chaal B.K., Gupta A.P., Wastuwidyaningtyas B.D., Luah Y-H., Bozdech Z., "Histone Deacetylases Play a Major Role in the Transcriptional Regulation of the Plasmodium falciparum Life Cycle", *PLoS Pathog*, 2010, Vol. 6, No.1.
- [7] Renom M.A., Stuart A., Fiser A., Sánchez R., Melo F. And Sali A., "Comparative protein structure modeling of genes and genomes," *Annu Rev Biophys Biomol Struct.*, Vol. 29, 2000, pp. 291-325.
- [8] Lovell S.C., Davis I.W., Arendall W.B., de Bakker P.I., Word J.M., Prisant M.G., Richardson J.S. and Richardson D.C., "Structure validation by Calpha geometry: phi,psi and Cbeta deviation," *Proteins.*, Vol. 15, No. 3, 2000, pp. 437-50.
- [9] Dundas J., et. al., "CASTp: computed atlas of surface topography of proteins with structural and topographical mapping of functionally annotated residues", *Nucleic Acids Res.*, Vol. 34, 2006, pp. 116-118. 10.
- [10] Pettersen E.F., Goddard T.D., Huang C.C., Couch G.S., Greenblatt D.M., Meng E.C. and Ferrin TE, "UCSF Chimera - A Visualization System for Exploratory Research and Analysis," *J Comput Chem.*, Vol. 25, 2004, pp. 1605-1612.